

TRITERPENES FROM *ILEX ROTUNDA* FRUITS

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(Received in revised form 12 October 1988)

Key Word Index—*Ilex rotunda*; Aquifoliaceae; pentacyclic triterpene; rotungenic acid; rotundioic acid; ursolic acid; rotundic acid, peduncloside.

Abstract—Two new triterpenes have been isolated from the fruits of *Ilex rotunda* along with three known triterpenoids ursolic acid, rotundic acid and peduncloside. Their structures were characterized as 3 β ,19 α ,24-trihydroxyurs-12-ene-28-oic acid and 3 β ,19 α -dihydroxyurs-12-ene-23,28-dioic acid by spectral and chemical means.

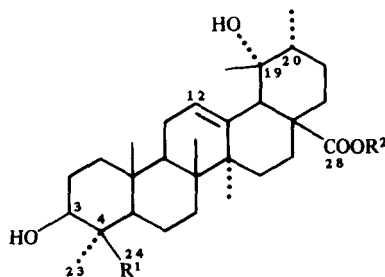
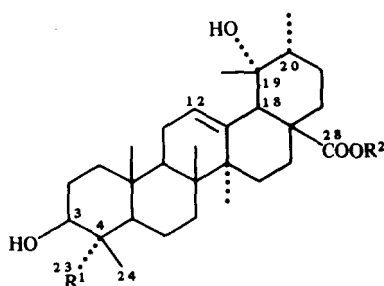
INTRODUCTION

There are some reports on the triterpenoids from *Ilex rotunda*. Rotundic acid (2) has been isolated from the seeds [1] and peduncloside (3) from the leaves [2]. *Ilex rotunda* yields red fruits in winter, from which we isolated two new ursene-type triterpenes rotungenic acid (4) and rotundioic acid (5) along with three known triterpenoids ursolic acid (1), rotundic acid (2) and its ester glucoside peduncloside (3). This paper describes the isolation and identification of these triterpenoids.

RESULTS AND DISCUSSION

The methanol extract of the fresh fruits afforded a mixture of triterpenoids which were separated by extensive chromatography to give ursolic acid (1), rotundic acid (2), peduncloside (28- β -glucopyranosyl rotundate) (3), and two new triterpenes 4 and 5.

Rotungenic acid (4), C₃₀H₄₈O₅, mp 295–298° (dec), exhibited the following spectral data. SIMS: m/z 489 [M+1]⁺; [α]_D + 50° (MeOH); UV 209 nm (ϵ 4000). Its IR spectrum showed the presence of hydroxyl (3600–3200 cm⁻¹) and carboxyl (1690 cm⁻¹) groups and double bond (1640 cm⁻¹). The ¹³C NMR spectrum revealed 30 carbon signals (Me-x6, -CH₂-x9, >CH-x4, -C-x5, -CH₂-O-x1, >CH-O-x1, -C-O-x1, >C=CH-x1, -CO-O-x1). Unequivocal information for the ring system and substitution mode in 4 was obtained from its EI mass spectrum. There were two characteristic peaks at m/z 264 (rel. int. 24%) and 223 (15%) denoting the retro-Diels-Alder cleavage fragments commonly found in the spectra of olean-12-ene or urs-12-ene derivatives possessing two hydroxyl groups in rings A/B and hydroxyl and carboxyl groups in rings D/E [1, 33, and three peaks at m/z 246 (44%), 219 (20%) and 201 (56%) indicated further successive losses of water and CO₂H



	R ¹	R ²		R ¹	R ²
2	CH ₂ OH	H	4	CH ₂ OH	H
3	CH ₂ OH	glu	6	CH ₂ OH	H 3 α -OH
5	COOH	H	7	COOH	H
8	CH ₂ OH	Me	9	CH ₂ OH	MC
10	COOMe	Me	11	CH ₂ OH	Me 3 α -OH
13	COOH	H 3 β -OAc	12	CH ₂ OAc	H 3 β -OAc
14	COOMe	Me 3 β -OAc	15	CHO	H
16	CHO	H	17	Me	H
18	CH ₂ OH	28-CH ₂ OH			

from the m/z 246 peak. Furthermore, there was a prominent peak at m/z 146 (74%) apparently due to the secondary retro-Diels-Alder cleavage of the m/z 218 fragment ion. These fragmentations resemble very closely those of rotundic acid (2). The ^{13}C NMR spectrum also indicated that it was closely related in structure to compound 2 except for the substitution mode of A ring (Table 1). The signal at δ 23.7 was assigned to the 4α -methyl group because it was observed at about δ 10 higher field than the 4β -methyl in most ursene and oleanane triterpenes possessing a 3β -hydroxy group [4–7]. Therefore, the hydroxymethyl group giving a signal at δ 64.6 must be present in the 4β -position.

A further proof on the substitution was obtained from the ^1H NMR spectrum of methyl rotungenate (9) (Table 2). The H-3 α signal (6 3.44, *dd*) showed the upfield shift of SO.16 compared to that of methyl rotundate (8), attributed to the loss of a shielding effect by the equatorial 4α -CH₂OH group. On the other hand, a W-type long range coupling was observed between the H-3 α signal and one (6 3.35) of the 4β -hydroxymethyl signals. These data indicated that compound 4 could be 19 α ,24-dihydroxyursolic acid, an epimer of barbinervic acid 6 [8], and the structure was also elucidated chemically as follows.

Rotungenic acid (4) was oxidized with chromic anhydride in pyridine to give an 24-aldehyde (15), followed by

Wolff-Kishner reduction to give 3 β ,19 α -dihydroxyurs-12-ene-28-oic acid (17), mp > 300° [9–11]. Therefore, the structure of rotungenic acid was established to be 3 β ,19 α ,24-trihydroxyurs-12-ene-28-oic acid (4).

Rotundioic acid (5), mp 295–298°, [α]_D + 50° (MeOH), revealed a molecular ion at m/z 502 [$\text{M}]^+$ corresponding to C₃₀H₄₆O₆ and retro-Diels-Alder fragmentation ions at m/z 264 and 238 from the cleavage of the C-ring of urs-12-ene derivative. The m/z 238 ion suggested the presence of one carboxyl group in rings A/B. This fragmentation and the IR and UV spectra (see Experimental) showed the presence of similar groups in 5 to those in 3 and 4 except for one additional carboxyl group instead of the hydroxymethyl group. Its ^{13}C NMR spectrum revealed thirty carbon signals including characteristic signals due to two carboxyl groups (6 178.2 and 178.3), a trisubstituted double bond (6 125.6 and 137.6) and two alcoholic carbons (6 70.4 and 73.3). It afforded a dimethyl acetate (14) which showed the presence of a 3β -acetoxyl group (6 5.17: 1H, *dd*, J = 11.5 and 4.5 Hz; H-3 α) in the ^1H NMR spectrum. A signal at δ 2.60 (1H, *br s*; H-1 8) suggested the presence of a 19-*O*-substituted urs-12-ene skeleton [12].

These facts indicated that compound 5 was related to 2 and 3 except for the exchanging of –CH₂OH group to –COOH. In the ^{13}C NMR spectrum of 5, the C-4 signal at δ 52.3 was shifted to downfield by ca 9 ppm from those of 2 or 3, and the chemical shift was very similar to that (653.0) of an oleanane triterpene gypsogenic acid [13] possessing an equatorial carboxyl group at C-23. On the other hand, the comparison of the spectrum with that of ilixgenin A (7) [14] having a C-24 axial carboxyl group revealed significant differences in the chemical shifts of the A and B ring carbons (Table 1). Therefore, one of the carboxyl groups in 5 should be restricted to C-23 and the structure 5 was also established as follows.

Oxidation of the C₂₃-aldehyde 16 derived from 2 with silver oxide gave rotundioic acid (5), while reduction of 5 and 8 with lithium aluminium hydride afforded the same tetraol 18. The spectral and chemical evidence elucidated the structure of 5 as 3 β ,19 α -dihydroxyurs-12-ene-23,28-dioic acid.

EXPERIMENTAL

Mps: uncorr. Concentrations were performed under red. pres. at bath temps not exceeding 50°. ^1H NMR spectra were obtained at 360 MHz and ^{13}C NMR spectra at 25.2 MHz. All the compounds were finally purified by HPLC on a C₁₈ semiprep column using H₂O–MeOH solvent system.

Plant material. The ripe fruits of the plant were collected at Kagoshima University in March 1980.

Extraction and isolation. The ripe fruits (2 kg) were extracted with MeOH (3 × 10 l). After concn to 500 ml, H₂O (500 ml) was added and the mixture extracted with Et₂O and EtOAc to give 15 and 1.2 g of extracts, respectively. The Et₂O extract (5 g) was fractionated by CC using a MeOH–CH₂Cl₂ solvent system to give six fractions, I (1% MeOH–CH₂Cl₂): 350 mg, II (2% MeOH–CH₂Cl₂): 2.5 g, III (3% MeOH–CH₂Cl₂): 600 mg, IV (5% MeOH–CH₂Cl₂): 600 mg, V (7% MeOH–CH₂Cl₂): 750 mg and VI (10% MeOH–CH₂Cl₂): 100 mg. Rechromatography of the fraction I afforded ursolic acid (1) (35 mg). After rechromatography, the fraction II (125 mg) was separated by repeated HPLC using 3040% H₂O/MeOH as the solvent to give 2 (80 mg), 4 (8 mg) and 5 (4.5 mg). HPLC separation of the fraction IV (60 mg) with 40–55% H₂O–MeOH afforded 3 (15 mg)

Table 1. ^{13}C NMR spectral data (δ) for compounds 2, 4, 5 and 7

c	2	4	5	7*
1	38.9	38.8	38.5	39.8
2	21.1	28.5	26.9	29.1
3	73.7	80.3	73.3	78.3
4	42.9	43.2	52.2	49.2
5	48.8	56.5	49.6	56.9
6	18.9	19.3	19.5	20.9
7	33.4	34.0	34.5	33.9
8	40.4	40.4	39.8	40.2
9	47.9	47.9	45.9	47.2
10	37.3	37.2	36.8	37.9
11	24.1	24.3	24.1	24.5
12	128.1	127.9	125.6	128.1
13	140.0	140.0	137.6	139.9
14	42.2	42.1	40.0	42.2
15	29.4	29.1	31.1	29.1
16	26.5	26.5	25.4	26.5
17	48.3	48.4	45.7	48.3
18	54.7	54.7	52.3	54.7
19	72.7	72.8	70.4	72.7
20	42.4	42.4	39.8	42.2
21	27.0	27.0	24.8	27.0
22	38.5	38.6	36.1	38.4
23	68.2	23.7	178.2	24.2
24	13.1	64.6	9.8	180.6
25	17.3	17.2	13.8	13.9
26	16.8	16.8	14.4	17.1
27	24.9	24.7	24.6	24.5
28	180.7	180.8	178.3	180.6
29	27.2	27.2	26.9	27.0
30	16.0	16.1	14.8	16.8

*Cited from ref. [10].
Measured in C₅D₅N.

Table 2. ^1H NMR spectral data for compounds **8–11** (CDCl_3 , 360 MHz)

	8			9			10			11*		
H	δ	Mult	$J(\text{Hz})$	δ	Mult	$J(\text{Hz})$	δ	Mult	$J(\text{Hz})$	δ	Mult	$J(\text{Hz})$
3	3.60	<i>dd</i>	10, 6	3.44	<i>br dd</i>	11.5, 4.5	4.00	<i>dd</i>	11, 4.5	3.83	<i>t</i>	3
12	5.35	<i>brt</i>	3.5	5.35	<i>brt</i>	3.5	5.35	<i>br t</i>	3.5	5.31	<i>m</i>	
18	2.59	brs		2.60	<i>brs</i>		2.60	<i>brs</i>		2.58	<i>s</i>	
23	3.39	<i>d</i>	10.5	1.26	<i>s</i>					0.89	<i>s</i>	
	3.68	<i>d</i>	10.5									
24	0.95	<i>s</i>		3.35	<i>brd</i>	11	1.15	<i>s</i>		3.49	<i>d</i>	11
				4.20	<i>d</i>	11				3.69	<i>d</i>	11
25	0.95	<i>s</i>		0.86	<i>s</i>		0.94	<i>s</i>		1.08	<i>s</i>	
26	0.68	<i>s</i>		0.66	<i>s</i>		0.68	<i>s</i>		0.66	<i>s</i>	
27	1.25	<i>s</i>		1.25	<i>s</i>		1.27	<i>s</i>		1.21	<i>s</i>	
29	1.21	<i>s</i>		1.25	<i>s</i>		1.22	<i>s</i>		1.25	<i>s</i>	
30	0.94	<i>d</i>	6	0.97	<i>d</i>	7	0.95	<i>d</i>	6.5	0.94	<i>d</i>	6
OMe	3.60	<i>s</i>		3.60	<i>s</i>		3.61	<i>s</i>		3.57	<i>s</i>	
							3.73	<i>s</i>				

*Cited from ref. [8].

and an unknown compound (1.5 mg). The EtOAc fraction (100 mg) also gave **3** (21 mg).

Rotungenic acid (4). Columns from H_2O –MeOH; mp 295–298° (d); $[\alpha]_{\text{D}} + 16^\circ$ (MeOH); SIMS m/z : 489 $[\text{M} + 1]^+$; EIMS m/z (rel. int.): 488 $[\text{M}]^+$ (4), 470 $[\text{M} - \text{H}_2\text{O}]^+$ (9), 452 $[\text{M} - 2\text{H}_2\text{O}]^+$ (9), 442 $[\text{M} - \text{HCOOH}]^+$ (16), 264 (24), 246 $[\text{264} - \text{H}_2\text{O}]^+$ (44), 223 (15), 219 (20), 218 (20), 205 $[\text{223} - \text{H}_2\text{O}]^+$ (30), 201 $[\text{264} - \text{COOH}]^+$ (56), 175 (55), 146 (74). IR $\nu_{\text{max}}^{\text{MeOH}}$ cm^{-1} : 3600–2600, 1690, 1640; UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (E): 209 (4000).

Rotundioic acid (5). Prisms from MeOH; mp 295–298° (d); $[\alpha]_{\text{D}} + 50^\circ$ (MeOH); SIMS m/z : 503 $[\text{M} + 1]^+$; EIMS m/z (rel. int.): 502 $[\text{M}]^+$ (4), 484 $[\text{M} - \text{H}_2\text{O}]^+$ (26), 456 $[\text{M} - \text{HCOOH}]^+$ (28), 264 (6), 246 (48), 238 (21), 219 (8), 218 (8), 201 (68), 175 (63), 146 (94); IR $\nu_{\text{max}}^{\text{MeOH}}$ cm^{-1} : 3600–3200, 1690, 1635; UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (E): 210 (4500); ^1H NMR (pyridine- d_5): δ 1.00, 1.09, 1.42, 1.60, 1.66 (each 3H, s), 1.12 (3H, d, $J = 7$ Hz), 3.02 (1H, s), 3.04 (1H, m), 4.61 (1H, dd, $J = 10$ and 6 Hz), 4.71 (1H, s, -OH), 5.59 (1H, br t).

Ursolic acid (1), Mp 290°; EIMS m/z : 456 $[\text{M}]^+$, 438, 410, 248 (base peak), 207, 203, 189.

Rotundic acid (2). Mp 272–274° (d); $[\alpha]_{\text{D}} + 24^\circ$ (MeOH); EIMS m/z : 488 $[\text{M}]^+$, 470, 452, 442, 264, 246, 223, 201, 175, 146; IR $\nu_{\text{max}}^{\text{MeOH}}$ cm^{-1} : 3500–3200, 1690, 1640.

Pedunculoidic acid (3) Mp 213–214°; $[\alpha]_{\text{D}} + 22^\circ$ (MeOH); SIMS m/z : 673 $[\text{M} + \text{Na}]^+$, 489 $[\text{651} - \text{C}_6\text{H}_{11}\text{O}_5]^+$, 471 $[\text{489} - \text{H}_2\text{O}]^+$, 425 $[\text{453} - \text{CO}]^+$, 407 $[\text{425} - \text{H}_2\text{O}]^+$; IR $\nu_{\text{max}}^{\text{MeOH}}$ cm^{-1} : 3500–3200, 1720.

Methyl rotungenate (9). Rotungenic acid (4) was treated with CH_2N_2 to give the methyl ester 10, mp 208–211° (d); $[\alpha]_{\text{D}} + 5.5^\circ$ (MeOH); EIMS m/z : 502 $[\text{M}]^+$; IR $\nu_{\text{max}}^{\text{MeOH}}$ cm^{-1} : 3500, 1725.

Diacetylrotungenic acid (12). Compound 4 was acetylated with Ac_2O in pyridine to give the diacetate (12), mp 164–167°; SIMS m/z : 573 $[\text{M} + 1]^+$; IR $\nu_{\text{max}}^{\text{MeOH}}$ cm^{-1} : 3500, 1735, 1690.

Dimethyl rotundioate (10). Rotundioic acid (5) was treated with CH_2N_2 to give the dimethyl ester 13; EIMS m/z : 530 $[\text{M}]^+$.

Acetylrotundioic acid (13). Compound 5 was acetylated with Ac_2O in pyridine to give the acetate 13, mp > 300°; SIMS m/z : 545 $[\text{M} + 1]^+$; IR $\nu_{\text{max}}^{\text{MeOH}}$ cm^{-1} : 3500, 1735, 1710, 1690.

Dimethyl acetylrotundioate (14). Compound 13 was methylated with CH_2N_2 to give the dimethyl ester 14, mp 210–212°; EIMS m/z : 572 $[\text{M}]^+$; ^1H NMR (CDCl_3): δ 0.67, 0.97, 1.19, 1.22, 1.26, 1.98 (each 3H, s), 0.95 (3H, d, $J = 7$ Hz), 2.60 (1H, br s), 3.60,

3.67 (each 3H, s), 5.17 (1H, dd, $J = 11.5$ and 4.5 Hz), 5.35 (1H, br t, $J = 3.5$ Hz).

Methyl rotundate (8). Mp 257–259° (d); $[\alpha]_{\text{D}} + 43^\circ$ (CHCl_3); EIMS m/z : 502 $[\text{M}]^+$.

Aldehyde 15. Compound 4 was oxidized with chromic anhydride in pyridine at room temp. to give 15, mp 200–202°; EIMS m/z : 486 $[\text{M}]^+$; ^1H NMR (CDCl_3): δ 0.76, 0.84, 1.20 (each 3H, s), 0.94 (3H, d, $J = 6.5$ Hz), 1.27 (6H, s), 2.53 (1H, s), 3.19 (1H, m), 5.34 (1H, br t), 9.76 (1H, s).

Aldehyde 16. Mp 205.5–207.5°; EIMS m/z : 486 $[\text{M}]^+$; ^1H NMR (pyridine- d_5): 69.61 (1H, s, -CHO).

Wolff–Kishner reduction of 15. Compound 15 was treated with hydrazine hydrate (64%) and KOH in triethyleneglycol at 195–200° for 6 hr. The product gave 17 (15%), mp > 300°; EIMS m/z : 472 $[\text{M}]^+$; ^1H NMR (CDCl_3 , + MeOH- d_4): δ 0.76, 0.78, 0.91, 0.99, 1.21, 1.26 (each 3H, s), 0.95 (3H, d, $J = 6.6$ Hz), 2.50 (1H, s), 3.22 (1H, dd, $J = 10$ and 5.5 Hz), 5.36 (1H, br t), which was also obtained from 16. Methyl ester; mp 123–125°.

Tetraol 18. Rotundioic acid (5) was treated with LiAlH_4 in THF to give a tetraol 18, mp 159–161°; SIMS m/z : 475 $[\text{M} + 1]^+$; ^1H NMR (CDCl_3): δ 0.91, 0.97, 0.99, 1.18, 1.30 (each 3H, s), 0.95 (3H, d, $J = 6$ Hz), 3.19 (1H, d, $J = 10.5$ Hz), 3.46 (2H, t, $J = 10$ Hz), 3.67 (1H, dd, $J = 9$ and 7 Hz), 3.74 (1H, d, $J = 10.5$ Hz), 5.26 (1H, br t), which was also obtained from 8. Triacetate; ^1H NMR (CDCl_3): δ 0.85, 0.95, 0.98, 1.00, 1.17, 1.29, 2.03, 2.05, 2.07 (each 3H, s), 2.36 (1H, dt, $J = 15$ and 18.5 Hz), 3.64, 4.00 (each 3H, d, $J = 11$ Hz), 3.72, 3.89 (each 1H, d, $J = 11.5$ Hz), 4.80 (1H, dd, $J = 5$ and 11 Hz), 5.26 (1H, br t).

Acknowledgement—This work was partially supported by Grants-in-Aid for the Scientific Research (No. 60540355) from the Ministry of Education, Science and Culture.

REFERENCES

- Oyama, T., Aoyama, H., Yamada, K., Mitsuhashi, T. and Sugiyama, N. (1968) *Tetrahedron Letters* 4639.
- Hase, T., Hagii, H., Ishizu, M., Ochi, M., Ichikawa, N. and Kubota, T. (1973) *Nippon Kagaku Kaishi* 778.
- Budjickiewicz, H., Wilson, J. M. and Djerassi, C. (1963) *J. Am. Chem. Soc.* 85, 3688.

4. Houghton, P. J. and Lian, L. M. (1986) *Phytochemistry* **25**, 1939.
5. Wenjuan, Q., Xiue, W., Junjie, Z., Fukuyama, Y., Yamada, T. and Nakagawa, K. (1986) *Phytochemistry* **25**, 913.
6. Ogiwara, K., Higa, M., Hobama, K. and Suga, T. (1987) *Phytochemistry* **26**, 783.
7. Chemli, R., Babadjamian, A., Fame, R., Boukef, K., Balansard, G. and Vidal, E. (1987) *Phytochemistry* **26**, 1785.
8. Takani, M., Kubota, K., Nozawa, M., Ushiki, T. and Takahashi, K. (1977) *Chem. Pharm. Bull.* **25**, 981.
9. Brieskorn, C. H. and Wunderer, H. (1967) *Chem. Ber.* **100**, 1252.
10. Bermejo, J., Beton, J. L., Fuente, G. de la and Gonzalez, A. G. (1967) *Tetrahedron Letters* **4649**.
11. Takani, T., Kawaguchi, S., Nishimura, K., Kubota, K., Tanabe, Y. and Takani, M. (1974) *Chem. Pharm. Bull.* **22**, 650.
12. Yoshioka, I., Sugawara, T. and Kitagawa, I. (1971) *Chem. Pharm. Bull.* **19**, 1700.
13. Oshima, Y., Ohsawa, T., Oikawa, K., Konno, C. and Hikino, H. (1984) *Planta Med.* **40**.
14. Hidaka, K., Ito, M., Matsuda, Y., Hohda, H., Yamazaki, K. and Yamahara, J. (1987) *Phytochemistry* **26**, 2023.